

Olerup SSP[®] KIR Genotyping

Product number:	104.101-12 – including <i>Taq</i> polymerase 104.101-12u – without <i>Taq</i> polymerase
Lot number:	3L9
Expiry date:	2024-06-01
Number of tests:	12
Number of wells per test:	27+1
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

This Product Description is only valid for Lot No. 3L9.

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

CHANGES COMPARED TO THE PREVIOUS OLERUP SSP[®] KIR GENOTYPING LOT (3K9)

The KIR Genotyping kit design, specificity and interpretation tables are based on IPD-KIR database 2.9.0.

The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
27	Moved, added	Moved, added	Primer pair added for 2DL2*004, negative control moved to mix 28
28	Added	Added	Negative control from mix 27

Changes in revision R01 compared to R00:

1. The sizes of the specific PCR products in wells 3, 5, 8 and 10 have been corrected.

Changes in revision R02 compared to R01:

1. The footnote referring to a lower yield of specific PCR product in primer mix 7 was modified.

Changes in R03 compared to R02:

1. Primer mix 14 amplifies the 3DS1*078 allele. The correction has been implemented in the specificity and interpretation tables.

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Lot-specific information

Well **28** contains Negative Control primer pairs, that will produce exon 4 and/or exon 5 amplicons for more than 97% of applicable KIR alleles as well as amplicons generated by positive control primer pairs constituent of all primer mixes in the Olerup SSP[®] product range.

PCR product sizes: 280bp KIR specific amplicons
 430bp Positive control

Length of PCR product	280	280	280	280
5'-primer¹	110	109	208	208
	5'-CAg ^{3'}	5'-CCT ^{3'}	5'-CCA ^{3'}	5'-CCg ^{3'}
3'-primer	187	187	288	288
	5'-ggT ^{3'}	5'-ggT ^{3'}	5'-gTC ^{3'}	5'-gTC ^{3'}
	187	187	288	288
	5'-ggT ^{3'}	5'-ggT ^{3'}	5'-ggT ^{3'}	5'-ggT ^{3'}
			288	288
			5'-gAT ^{3'}	5'-gAT ^{3'}
2DL1*	+		+	
2DL2*	+		+	
2DL3*	+		+	
2DL4*	N/A	N/A		+
2DL5A*	N/A	N/A	+	
2DL5B*	N/A	N/A	+	
2DS1*	+		+	
2DS2*	+		+	
2DS3*	+		+	
2DS4*		+	+	
2DS5*	+		+	
3DL1*	+		+	
3DL2*	+		+	
3DL3*	+		+	
3DS1*	+		+	
2DP1*	+		+	
3DP1*	+		+	

¹The codon position for KIR genes, in the 4th or 5th exon, matching the specificity-determining 3'-end of the primer is given. Codon numbering as on the www.ebi.ac.uk/ipd/kir web site. The sequence of the 3 terminal nucleotides of the primer is given.

PRODUCT DESCRIPTION

KIR Genotyping SSP typing

CONTENT

The primer set contains 5'- and 3'-primers for KIR Genotyping.

PLATE LAYOUT

Each test consists of 28 PCR reactions in a 32 well cut PCR plate. Wells 29 to 32 are empty.

1	2	3	4	5	6	7	8
9	10	11	12	13	14	15	16
17	18	19	20	21	22	23	24
25	26	27	NC	empty	empty	empty	empty

The 32 well cut PCR plate is marked with ‘KIR GENOTYP’ in silver/gray ink.

Well No. 1 is marked with the Lot No. ‘3L9’.

Wells 1 to 27 – KIR Genotyping primers.

Well 28 – Negative Control.

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.

The PCR plates are covered with a PCR-compatible foil.

Please note: When removing each 32 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

UNIQUELY IDENTIFIED ALLELES

All the KIR alleles available in the IPD KIR Sequence Database in December 2019¹ will be amplified by the primers in the KIR Genotyping SSP kit.

¹KIR alleles listed on the IPD KIR web page 2019-December-11, release 2.9.0, www.ebi.ac.uk/ipd/kir.

PROTOCOL

DNA EXTRACTION

Extracted, highly pure DNA is needed for SSP typings. DNA samples to be used for PCR-SSP HLA typing should be re-suspended in dH₂O. The A260/A280 ratio should be 1.6 – 2.0 by UV spectrophotometry for optimal band visualization during electrophoresis.

We recommend automated DNA extraction with the QIAGEN EZ1 DSP DNA Blood System. ACD blood should be used as starting material.

Alternatively, the DNA can be extracted by any preferred method yielding pure DNA. When using alternative methods, the DNA concentration should be adjusted to 30 ng/μl. **Do not use heparinised blood with these methods.**

Recommended DNA concentration using:
EZ1-extracted DNA, 15 ng/μl.
DNA extracted by other methods, 30 ng/μl.

Concentrations exceeding 50 ng/μl will increase the risk for nonspecific amplifications and weak extra bands. If necessary, dilute the extracted DNA in dH₂O.

DNA samples should not be re-suspended in solutions containing chelating agents such as EDTA, above 0.5 mM in concentration.

DNA samples may be used immediately after extraction or stored at +4°C for up to 2 weeks with no adverse effects on results. DNA samples can be stored at -20°C or colder for 9 months. The purity and concentration of extracted DNA samples that have been stored for a prolonged period should be tested for acceptability prior to HLA typing.

DNA samples should be shipped at +4°C or colder to preserve their integrity during transport.

PCR AMPLIFICATION

INSTRUMENT REQUIREMENTS

A thermocycler with the following minimum specifications should be used:

- heated lid with a temperature of 104°C for oil-free operation
- sample block (aluminum, silver, or gold-plated silver) for use with either a 96-well PCR plate or 0.2 ml thin-walled reaction tubes
- Olerup SSP kits are validated on the following cyclers.

Recommended ramp rates:

- GeneAmp 9700: GeneAmp 9700 cycler set to the 9600 mode. This correspond to a **sample ramp rate** of 1.6°C/s up and 0.8°C/s down.

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- ProFlex 1x96-well block: ProFlex PCR cycler with a block ramp rate of 3.0°C/s (each step 3.0°C/s). A **block ramp rate** of 3.0°C/s correspond to a sample ramp rate of 1.52°C/s up and 1.36°C/s down.
- ProFlex 2x96-well block: ProFlex PCR cycler with a block ramp rate of 3.0°C/s (each step 3.0°C/s). A **block ramp rate** of 3.0°C/s correspond to a sample ramp rate of 1.9°C/s up and 1.6°C/s down.

Note: Higher ramp rates than the equivalent to the described may have an effect on the typing results. Please also note that the effect on the typing may differ between different non-validated cyclers depending on the settings.

- temperature range of 4.0°C to 99.9°C
- temperature accuracy of ±0.25°C over the range of 35°C to 99.9°C
- sample block temperature uniformity of ≤0.75°C over the range of 55°C to 95°C
- temperature calibration traceable to a reference standard (i.e., NIST)

Program the thermocycler using the PCR Cycling Parameters specified below.

For specific thermocycler information refer to the manufacturer’s user manual. Thermocyclers should be calibrated according to ASHI (American Society of Histocompatibility and Immunogenetics) or EFI (European Federation of Immunogenetics) accreditation rules.

Program the thermocycler before starting the Directions for Use described below.

104.101-12 – including Taq polymerase

For one KIR Genotyping typing, begin by adding to well No. 28, i.e. the well with the negative control primer pairs:

7 µl dH₂O

3 µl PCR Master Mix complete with Taq,

then add at room temperature in a 0.5 ml tube:

32 x 2 µl = 64 µl DNA (30 ng/µl)

32 x 3 µl = 96 µl PCR Master Mix complete with Taq – mix well

before taking your aliquot

32 x 5 µl = 160 µl dH₂O

Mix well, dispense 10 µl of the DNA-PCR Master Mix-H₂O mixture into each of the 27 wells of an KIR Genotyping typing, i.e. wells 1 to 27. Cover the primer tray(s) with the provided adhesive seals. Check that all reaction wells are completely covered to prevent evaporative loss during PCR amplification.

104.101-12u – without Taq polymerase

For one KIR Genotyping typing, begin by adding at room temperature in a 0.5 ml tube:

32 x 3 µl = 96 µl PCR Master Mix without Taq – mix well before taking your aliquot

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2.5 µl *Taq* polymerase (5 units/µl)

Mix well, dispense 3 µl of the PCR Master Mix-*Taq* mixture from the 0.5 ml tube into well No. 28, i.e. the well with the negative control primer pairs. Then add 7 µl dH₂O to well 28.

Then add at room temperature to the 0.5 ml tube containing 96 + 2.5 - 3 = 95.5 µl PCR Master Mix-*Taq* mixture:

32 x 2 µl = 64 µl DNA (30 ng/µl)

32 x 5 µl - 2.5 µl = 157.5 + 5 µl dH₂O

Mix well, dispense 10 µl of the DNA-PCR Master Mix-*Taq*-H₂O mixture into each of the 26 wells of an KIR Genotyping typing, i.e. wells 1 to 26. Cover the primer tray(s) with the provided adhesive seals. Check that all reaction wells are completely covered to prevent evaporative loss during PCR amplification.

Use a 96 well thermal cycler with a heated lid. The temperature gradient across the heating block should be < 1°C.

PCR cycling parameters:

1. 1 cycle	94°C	2 min	denaturation
2. 10 cycles	94°C	10 sec.	denaturation
	65°C	60 sec.	annealing and extension
3. 20 cycles	94°C	10 sec.	denaturation
	61°C	50 sec.	annealing
	72°C	30 sec.	extension
4. End - hold	RT		if less than 8 hours
	4°C		if longer than 8 hours

Total reaction volume in each well, 10 µl.

The same PCR cycling parameters are used for all the Olerup SSP kits.

AGAROSE GEL ELECTROPHORESIS

Prepare a 2% (w/v) agarose gel in 0.5 x TBE buffer. Dissolve the agarose by boiling in a microwave oven. Let the gel solution cool to 60°C. Stain the gel prior to casting with ethidium bromide (10 mg/ml), 5 µl per 100 ml gel solution. For maximal ease of handling use our ethidium bromide dropper bottles (Product No. 103.301-10), 1 drop of ethidium bromide solution per 50-75 ml of gel. **Note: Ethidium bromide is a carcinogen. Handle with appropriate personal protective equipment.**

Load the PCR products, preferably using an 8-channel pipette. Load a DNA size marker (100 base pair ladder, Product No. 103.201-100) in one well per row.

Run the gel in 0.5 x TBE buffer, without re-circulation of the buffer, for 15-20 minutes at 8-10 V/cm.

DOCUMENTATION AND INTERPRETATION

Put the gel on a UV transilluminator and document by photography.

Record the presence and absence of specific PCR products. The relative lengths of the specific PCR products are helpful in the interpretation of the results.

Record the presence and relative lengths of the internal positive control bands. The differently sized control bands will help in the correct orientation of the typing as well as in kit identification.

Lanes without either control band or specific PCR products should be repeated.

Interpret the typings with the ***lot-specific Interpretation and Specificity Tables***.

PCR MASTER MIXES

The PCR Master Mix complete with *Taq* polymerase contains:

<i>Taq</i> polymerase	0.4 unit per 10 µl SSP reaction
nucleotides	final concentration of each dNTP is 200 µM
PCR buffer	final concentrations: 50 mM KCl, 1.5 mM MgCl ₂ , 10 mM Tris-HCl pH 8.3, 0.001% w/v gelatin
glycerol	final concentration of glycerol is 5%
cresol red	final concentration of cresol red is 100 µg/ml

The same PCR Master Mix complete with *Taq* is used for all Olerup SSP kits.

The PCR Master Mix without *Taq* polymerase contains:

nucleotides	final concentration of each dNTP is 200 µM
PCR buffer	final concentrations: 50 mM KCl, 1.5 mM MgCl ₂ , 10 mM Tris-HCl pH 8.3, 0.001% w/v gelatin
glycerol	final concentration of glycerol is 5%
cresol red	final concentration of cresol red is 100 µg/ml

The same PCR Master Mix without *Taq* is used for all Olerup SSP kits.

SPECIFICITY TABLE

KIR Genotyping SSP typing

Specificities and sizes of the PCR products of the 27+1 primer mixes used for KIR SSP Genotyping SSP.

Primer Mix	Size of spec. PCR product ¹	Size of control band ²	KIR Gene	Amplified KIR ³ alleles
1	145 bp	800 bp	2DL1	0010101-038
2^{4,7,9}	65 bp	1070 bp	2DL2	004, 011
	150 bp		2DL2	0010101-015
	225 bp		2DL2	004, 011
3^{4,7}	130 bp	1070 bp	2DL3	0010101 ^w -0020103 ^w , 004 ^w -015 ^w , 017 ^w -036 ^w
	520 bp		2DL3	0010101-011, 013-026, 028-036
4	200 bp	1070 bp	2DL4	00101-054
5⁶	140 bp	1070 bp	2DL5A	0010101-00105, 0050101-0050104, 01201-01202, 014-015, 021-024
			2DL5B	0020101-0020104, 0020106-004, 00601-011, 01301-01304, 016-020
6⁵	1650 bp	430 bp	2DL5A	0010101-00105, 0050101-0050104, 01201-01202, 014-015, 021-024
7^{5,6,7,8}	1650 bp	515 bp	2DL5B	0020101-0020104, 0020106-004, 00601-011, 01301-01304, 016-020
8⁴	105 bp	1070 bp	2DS1	001-006, 008-009
9	205 bp	1070 bp	2DS2	0010101-009
10	140 bp	1070 bp	2DS3	00101-008
11	215 bp	1070 bp	2DS4	0010101-00106, 01101-01102, 014-017, 019-020
12	200 bp	1070 bp	2DS4	0030101-0030104, 0040101-0040102, 0060101-0060102, 007-010, 012, 013, 018
13^{4,7}	110 bp	1070 bp	2DS5	001-017
14	135 bp	1070 bp	3DL1	0010101-0020105, 0040101-00404, 0050101-0090104, 0150101-044, 051-054, 056, 057, 05901-077, 079-081N, 086-103, 109-1190102
			3DS1	078
			3DL2	0010101-115
15	200 bp	1070 bp	3DL2	0010101-115
16⁴	115 bp	1070 bp	3DL3	00101-094
17	130 bp	1070 bp	3DS1	010-014, 045-050, 055, 058, 078, 082-085, 104-108
18	165 bp	1070 bp	2DP1	00101-004, 006-024

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Lot-specific information

19⁴	125 bp	1070 bp	3DP1	001-015
20	235 bp	1070 bp	3DP1	0030101-0030402, 005, 006, 008, 01001-1002, 013-015
21	145 bp	1070 bp	2DS1	001
22^{4,10}	95 bp 235 bp	1070 bp	2DS1 3DP1	0020101-009 001-002, 004, 007, 0090101-00902, 011-012
23	210 bp	1070 bp	3DL1	0040101-00404, 019, 021, 036, 037, 039, 056, 072, 091, 110, 117
24^{4,7,8}	100 bp	1070 bp	2DL4	00101-00603, 010, 01201-01202, 014-016, 018, 021-026, 028-034, 036-042, 045, 049, 050, 054
25	195 bp	1070 bp	2DL5B 3DP1	0020101, 0020102 [?] , 0020103, 0020104 [?] , 0020106 [?] , 0020107-0020108, 0020201 [?] , 00203 [?] -00204 [?] , 004, 00601, 00603 [?] , 0070101, 0070102 [?] , 0080101, 0080102 [?] , 00802, 00803 [?] -009 [?] , 010-011, 01301 [?] -01302 [?] , 01304 [?] , 016 [?] -020 [?] 001, 002 [?] , 007, 0090101-0090103, 01002 [?] -015 [?]
26	160 bp	1070 bp	2DL5A 2DS5 3DP1	0010101-00105, 0050101, 0050103-0050104, 01201-01202, 014-015 [?] , 021-024 [?] 001 [?] , 0020101-0020104, 00202 [?] -00502 [?] , 00801 [?] -009 [?] , 011 [?] -017 [?] 004
27	180 bp	1070 bp	2DL2	004
28^{6,11}	-	-	-	Negative control

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of KIR SSP typings.

When the primers in a primer mix can give rise to KIR-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene.

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Lot-specific information

The internal positive control bands are 1070, 800, 515 or 430 base pairs respectively, well distribution as outlined in the table. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

³KIR alleles listed on the IPD KIR web page 2019-December-11, release 2.9.0, www.ebi.ac.uk/ipd/kir.

⁴Specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

⁵The specific PCR product generated by primer mixes 6 and 7 are longer than the internal positive control band and the positive control band may be weaker than for other KIR primer mixes.

⁶Primer mixes 5, 7 and 28 have a tendency to give rise to primer oligomer formation.

⁷Primer mixes 2, 3, 7, 13 and 24 may have tendencies of unspecific amplifications.

⁸Primer mixes 7 and 24 may give rise to a lower yield of specific PCR product than the other KIR primer mixes, most pronounced for primer mix 7.

⁹The 2DL2*004 and 2DL2*011 and the 2DL2*0010101-010 and 012-013 alleles may be distinguished by the different sizes of the specific PCR product in primer mix 2; three specific PCR fragments of 65, 150 and 225 bp in the 2DL2*004 and 2DL2*011 alleles and one specific PCR fragment of 150 bp in the 2DL2*0010101-00304, 005-010 and 012-013 alleles.

¹⁰The 2DS1 and the 3DP1 amplicons in primer mix 22 may be distinguished by the different sizes of the specific PCR product; a specific PCR fragment of 95 bp for the 2DS1*0020101-009 alleles and a specific PCR fragment of 235 bp for the 3DP1*001-002, 004, 007, 0090101-00902 and 011-012 alleles.

¹¹Well 28 contains negative control primer pairs, that will produce exon 4 and/or exon 5 amplicons for more than 97% of applicable KIR alleles as well as amplicons generated by positive control primer pairs.

Abbreviations

'w', might be weakly amplified.

'?', nucleotide sequence information not available for the primer matching sequence.

PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11	12	13	14
Length of spec. PCR product	145	65 150 225	130 520	200	140	1650	1650	105	205	140	215	200	110	135
Length of int. pos. control ¹	800	1070	1070	1070	1070	430	515	1070	1070	1070	1070	1070	1070	1070
5'-primer(s) ²	130 5'-gAA ^{3'}	156 5'-AAA ^{3'}	332 5'-TCg ^{3'}	208 5'-CCg ^{3'}	226 5'-CCA ^{3'}	-16 5'-TCA ^{3'}	-16 5'-TCg ^{3'}	165 5'-gAg ^{3'}	140 5'-gTA ^{3'}	236 5'-CAC ^{3'}	229 5'-CTA ^{3'}	234 5'-TCT ^{3'}	142 5'-ACC ^{3'}	136 5'-CAA ^{3'}
	130 5'-TAA ^{3'}	208 5'-CCA ^{3'}	344 5'-CTg ^{3'}				-16 5'-Tgg ^{3'}	165 5'-gAA ^{3'}						208 5'-CCA ^{3'}
		262 5'-ggA ^{3'}	344 5'-CTg ^{3'}											166 5'-CAA ^{3'}
			378 5'-TAT ^{3'}											238 5'-CCg ^{3'}
3'-primer(s) ³	165 5'-gCg ^{3'}	195 5'-ATg ^{3'}	350 5'-CAA ^{3'}	262 5'-ggA ^{3'}	276 5'-gAg ^{3'}	27 5'-ACA ^{3'}	27 5'-ACA ^{3'}	185 5'-gAC ^{3'}	195 5'-ATg ^{3'}	266 5'-CCT ^{3'}	288 5'-ggA ^{3'}	288 5'-ggA ^{3'}	165 5'-gTg ^{3'}	
		243 5'-ACA ^{3'}	351 5'-ACC ^{3'}											
		269 5'-TAC ^{3'}	405 5'-CgA ^{3'}											
Well No.	1	2	3	4	5	6	7	8	9	10	11	12	13	14

Well No.	15	16	17	18	19	20	21	22	23	24	25	26	27
Length of spec. PCR product	200	115	130	165	125	235	145	95 235	210	100	195	160	180
Length of int. pos. control ¹	1070	1070	1070	1070	1070	1070	1070	1070	1070	1070	1070	1070	1070
5'-primer(s) ²	110 5'-ggg ^{3'}	156 5'-CCC ^{3'}	133 5'-TCT ^{3'}	29 5'-CAT ^{3'}	25 5'-Tgg ^{3'}	2 nd I 5'-gCC ^{3'}	130 5'-gAA ^{3'}	130 5'-gAA ^{3'}	31 5'-TCA ^{3'}	324 5'-gTA ^{3'}	up ⁴ 5'-AAG ^{3'}	up ⁵ 5'-CCg ^{3'}	106 5'-CAC ^{3'}
								2 nd I 5'-TCC ^{3'}	31 5'-TCA ^{3'}				
3'-primer(s) ³	164 5'-CAA ^{3'}	181 5'-gTA ^{3'}	163 5'-ggA ^{3'}	71 5'-TAC ^{3'}	54 5'-TAC ^{3'}	54 5'-TAC ^{3'}	165 5'-gCC ^{3'}	54 5'-TAC ^{3'}	86 5'-CCA ^{3'}	344 5'-Tgg ^{3'}	1 st I 5'-TCA ^{3'}	-16 5'-gAT ^{3'}	165 5'-gCg ^{3'}
								165 5'-gCT ^{3'}					
Well No.	15	16	17	18	19	20	21	22	23	24	25	26	27

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 1070, 800, 515 or 430 base pairs respectively, well distribution as outlined in the table. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

²The codon position matching the specificity-determining 3'-end of the primer is given. Codon numbering as on the KIR web page 2018-November-30, release 2.8.0, www.ebi.ac.uk/ipd/kir. The sequence of the 3 terminal nucleotides of the primer is given.

³The codon position matching the specificity-determining 3'-end of the primer. Codon numbering as on the KIR web page 2018-November-30, release 2.8.0, www.ebi.ac.uk/ipd/kir. The sequence of the 3 terminal nucleotides of the primer is given in the anti-sense direction.

⁴Primer located upstream of the 1st exon, 84 nucleotides upstream of codon -21.
⁵Primer located upstream of the 1st exon, 104 nucleotides upstream of codon -21.

		CELL LINE VALIDATION SHEET															
		KIR Genotyping primer set ²															
		Well															
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Prod. No.:		201787601	201787602	202022403	201787604	201787605	201787606	202022407	201787608	201787609	201787610	201787611	201787612	201787613	201787614	201787615	201787616
IHCW cell line ¹																	
1	9001 SA	+	-	+	+	-	-	-	-	-	-	+	-	-	+	+	+
2	9280 LK707	-	+	-	+	+	-	+	+	+	-	-	+	+	+	+	+
3	9011 E4181324	+	-	+	+	-	-	-	-	-	-	+	+	-	-	+	+
4	9275 GU373	+	-	+	+	-	-	-	-	-	-	+	+	-	+	+	+
5	9009 KAS011	+	-	+	+	+	+	-	+	-	-	-	+	+	+	+	+
6	9353 SM	+	-	+	+	+	+	-	+	-	+	+	-	-	+	+	+
7	9020 QBL	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+
8	9025 DEU	+	+	+	+	-	-	-	-	+	-	-	+	-	+	+	+
9	9026 YAR	+	-	+	+	-	-	-	-	-	-	+	+	-	+	+	+
10	9107 LKT3	+	-	+	+	-	-	-	-	-	-	+	+	-	+	+	+
11	9051 PITOUT	+	+	+	+	-	-	-	-	+	-	-	+	-	+	+	+
12	9052 DBB	+	+	+	+	+	-	+	-	+	+	-	+	-	+	+	+
13	9025 JESTHOM	+	+	-	+	+	-	+	-	+	+	+	+	-	+	+	+
14	9071 OLGA	+	-	+	+	+	+	-	+	-	-	-	+	+	+	+	+
15	9075 DKB	+	-	+	+	-	-	-	-	-	-	+	-	-	+	+	+
16	9037 SWEIG007	+	-	+	+	-	-	-	-	-	-	-	+	-	+	+	+
17	9282 CTM3953540	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+
18	9257 32367	+	-	+	+	-	-	-	-	-	-	-	+	-	+	+	+
19	9038 BM16	+	-	+	+	-	-	-	-	-	-	-	+	-	+	+	+
20	9059 SLE005	+	-	+	+	-	-	-	-	-	-	+	+	-	+	+	+
21	9064 AMALA	+	+	+	+	+	+	-	+	+	-	+	-	+	+	+	+
22	9056 KOSE	+	+	+	+	-	-	-	-	+	-	-	+	-	+	+	+
23	9124 IHL	+	+	+	+	-	-	-	-	+	-	+	+	-	+	+	+
24	9035 JBUSH	+	-	+	+	-	-	-	-	-	-	+	+	-	+	+	+
25	9049 IBW9	+	-	+	+	-	-	-	-	-	-	+	+	-	+	+	+
26	9285 WT49	+	+	+	+	+	-	+	+	+	-	+	+	+	+	+	+
27	9191 CH1007	+	+	+	+	+	-	+	-	+	+	+	+	-	+	+	+
28	9320 BEL5GB	+	+	-	+	+	-	+	-	+	+	-	+	-	+	+	+
29	9050 MOU	+	-	+	+	-	-	-	-	-	-	-	+	-	+	+	+
30	9021 RSH	+	+	+	+	+	-	+	-	+	-	+	+	+	+	+	+
31	9019 DUCAF	+	-	+	+	-	-	-	-	-	-	-	+	-	+	+	+
32	9297 HAG	+	-	+	+	-	-	-	-	-	-	+	+	-	+	+	+
33	9098 MT14B	+	-	+	+	-	-	-	-	-	-	+	-	-	+	+	+
34	9104 DHIF	+	+	+	+	-	-	-	-	+	-	+	+	-	+	+	+
35	9302 SSTO	+	+	+	+	-	-	-	-	+	-	+	+	-	+	+	+
36	9024 KT17	+	-	+	+	+	+	-	+	-	+	-	+	-	+	+	+
37	9065 HHKB	+	+	+	+	+	+	-	+	+	-	-	+	+	+	+	+
38	9099 LZL	+	+	+	+	+	-	+	+	+	+	-	-	+	+	+	+
39	9315 CML	+	+	-	+	+	+	+	+	+	+	-	+	-	+	+	+
40	9134 WHONP199	+	-	+	+	-	-	-	-	-	-	+	-	-	+	+	+
41	9055 H0301	+	+	-	+	+	-	+	-	+	+	-	-	-	+	+	+
42	9066 TAB089	+	-	+	+	-	-	-	-	-	-	-	+	-	+	+	+
43	9076 T7526	+	-	+	+	+	+	-	+	-	-	+	-	+	+	+	+
44	9057 TEM	+	+	+	+	+	-	+	-	+	+	-	+	-	+	+	+
45	9239 SHJO	+	+	+	+	+	-	+	-	+	-	+	-	+	+	+	+
46	9013 SCHU	+	-	+	+	-	-	-	-	-	-	+	+	-	+	+	+
47	9045 TUBO	+	-	+	+	-	-	-	-	-	-	-	+	-	+	+	+
48	9303 TER-ND	+	-	+	+	-	-	-	-	-	-	-	+	-	+	+	+



CELL LINE VALIDATION SHEET														
KIR Genotyping primer set ²														
			Well											
			17	18	19	20	21	22	23	24	25	26	27	
			Prod. No.:	201787617	201787618	201787619	201787620	201787621	201787622	201787623	201787624	201787625	201787626	202022427
IHC cell line ¹														
1	9001	SA	-	+	+	+	-	-	-	+	-	-	-	
2	9280	LK707	-	-	+	-	-	+	-	-	+	+	-	
3	9011	E4181324	+	+	+	+	-	+	+	+	-	+	-	
4	9275	GU373	-	+	+	+	-	-	-	+	-	-	-	
5	9009	KAS011	+	+	+	+	-	+	-	+	-	+	-	
6	9353	SM	+	+	+	+	-	+	-	+	-	+	-	
7	9020	QBL	+	+	+	+	-	+	+	+	+	+	-	
8	9025	DEU	-	+	+	+	-	+	-	-	+	-	-	
9	9026	YAR	-	+	+	+	-	-	-	+	-	-	-	
10	9107	LKT3	-	+	+	+	-	-	-	+	-	-	-	
11	9051	PITOUT	-	+	+	+	-	+	+	-	+	-	-	
12	9052	DBB	-	+	+	+	-	-	+	-	+	-	-	
13	9025	JESTHOM	-	+	+	+	-	+	-	+	+	-	-	
14	9071	OLGA	+	+	+	+	-	+	-	+	-	+	-	
15	9075	DKB	-	+	+	+	-	-	-	+	-	-	-	
16	9037	SWEIG007	-	+	+	+	-	-	+	-	-	-	-	
17	9282	CTM3953540	+	+	+	+	-	+	+	+	+	+	-	
18	9257	32367	-	+	+	+	-	-	+	-	-	-	-	
19	9038	BM16	-	+	+	+	-	-	-	-	-	-	-	
20	9059	SLE005	-	+	+	+	-	-	-	+	-	-	-	
21	9064	AMALA	+	+	+	+	-	+	-	+	+	+	-	
22	9056	KOSE	-	+	+	+	-	+	+	-	+	-	-	
23	9124	IHL	-	+	+	+	-	+	+	+	+	-	-	
24	9035	JBUSH	-	+	+	+	-	-	-	+	-	-	-	
25	9049	IBW9	-	+	+	+	-	-	+	+	-	-	-	
26	9285	WT49	-	+	+	+	-	+	-	+	+	+	-	
27	9191	CH1007	-	+	+	+	-	-	-	+	+	-	-	
28	9320	BEL5GB	-	+	+	+	-	+	+	+	+	-	-	
29	9050	MOU	-	+	+	+	-	-	+	-	-	-	-	
30	9021	RSH	-	+	+	+	-	-	-	+	+	-	-	
31	9019	DUCAF	-	+	+	+	-	-	-	-	-	-	-	
32	9297	HAG	-	+	+	+	-	-	-	+	-	-	-	
33	9098	MT14B	-	+	+	+	-	-	-	+	-	-	-	
34	9104	DHIF	-	+	+	+	-	+	-	+	+	-	-	
35	9302	SSTO	-	+	+	+	-	+	-	+	+	-	-	
36	9024	KT17	+	+	+	+	-	+	-	+	-	+	-	
37	9065	HHKB	+	+	+	+	-	+	-	+	+	+	-	
38	9099	LZL	-	+	+	+	-	+	-	+	+	-	-	
39	9315	CML	+	+	+	+	-	+	+	+	+	+	-	
40	9134	WHONP199	-	+	+	+	-	-	-	+	-	-	-	
41	9055	H0301	-	+	+	+	-	+	-	+	+	-	-	
42	9066	TAB089	-	+	+	+	-	-	-	-	-	-	-	
43	9076	T7526	+	+	+	+	-	+	-	+	-	+	-	
44	9057	TEM	-	+	+	+	-	-	-	+	+	-	-	
45	9239	SHJO	-	+	+	+	-	-	-	+	+	-	-	
46	9013	SCHU	-	+	+	+	-	-	-	+	-	-	-	
47	9045	TUBO	-	+	+	+	-	-	-	-	-	-	-	
48	9303	TER-ND	-	+	+	+	-	-	-	-	-	-	-	

Lot No.: 3L9

Lot-specific information

¹The provided cell line KIR specificities are retrieved from the www.ebi.ac.uk/ipd/kir web site. The specificity of an individual cell line may thus be subject to change.

²The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solution 21 were available.

One additional primer in primer solution 3 was tested by separately adding another 5'-primer.

In primer solution 21 it was only possible to test the 5'-primer, the 3'-primer was not possible to test.

In primer solutions 1, 2, 3, 7 and 16 one of the 5'-primers were not possible to test, and in primer mix 2, 16 and 24 one 3'-primer was not possible to test.

Lot No.: 3L9

Lot-specific information

TRADEMARKS USED IN THIS DOCUMENT/PRODUCT

Olerup SSP[®] is a registered trademark of *CareDx* AB.
 Qiagen[™] is a trademark of QIAGEN.

WARRANTY

CareDx AB warrants its products to the original purchaser against defects in materials and workmanship under normal use and application. *CareDx* AB’s sole obligation under this warranty shall be to replace, at no charge, any product that does not meet the performance standards stated on the product specification sheet.

This warranty applies only to products that have been handled and stored in accordance with *CareDx* AB’s recommendations, and does not apply to products that have been the subject of alternation, misuse, or abuse.

All claims under this warranty must be directed to *CareDx* AB in writing and must be accompanied by a copy of the purchaser’s invoice. This warranty is in lieu of all other warranties, expressed or implied, including the warranties of merchantability and fitness for a particular purpose. In no case shall *CareDx* AB be liable for incidental or consequential damages.

This product may not be reformulated, repacked or resold in any form without the written consent of *CareDx* AB, Franzengatan 5, SE-112 51 Stockholm, Sweden. Handle all samples as if capable of transmitting disease. All work should be performed wearing gloves and appropriate protection.

GUARANTEE

CareDx AB guarantees that the primers in the *Olerup* SSP[®] typing trays have the specificities given in the lot-specific Specificity and Interpretation Tables of the product insert.

When stored at –20°C, the dried primers are stable for 48 months from the date of manufacture.

When stored at –20°C, the PCR Master Mix including *Taq* polymerase and the PCR Master Mix without *Taq* polymerase are stable for 51 months from the date of manufacture.

KIR Genotyping
104.101-12 – including Taq polymerase
104.101-12u – without Taq polymerase

Product Insert

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Visit www.caredx.com for
“Instructions for Use” (IFU)

Lot No.: 3L9

Lot-specific information

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